Ultrasmall InGa(As)P Dielectric and Plasmonic Nanolasers

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ABSTRACT: Nanolasers have great potential for both on-chip light sources and optical barcoding particles. We demonstrate ultrasmall InGaP and InGaAsP disk lasers with diameters down to 360 nm (198 nm in height) in the red spectral range. Optically pumped, room-temperature, single-mode lasing was achieved from both disk-on-pillar and isolated particles. When isolated disks were placed on gold, plasmon polariton lasing was obtained with Purcell-enhanced stimulated emission. UV lithography and plasma ashing enabled wafer-scale fabrication of nanodisks with an intended random size variation. Silica-coated nanodisk particles generated stable subnanometer spectra from within biological cells across an 80 nm bandwidth from 635 to 715 nm.

KEYWORDS: nanolaser, lasing, nanophotonics, plasmonic, cell barcoding

Submicron- and nanoscale lasers have received interest for basic research and applications including optical communications, integrated photonics, chemical sensing, and biological probing. A small device volume is the defining characteristic of nanolasers. The small size not only provides some intriguing physical phenomena such as large Purcell effects and threshold-less lasing, but is also important for practical applications. A small volume of the gain medium is beneficial for low pump energy consumption. Also importantly, smaller sizes are generally preferred for biological applications to facilitate cellular uptake or attachment on the cell membrane with minimal physical perturbations on cells. Furthermore, a small size is critical for the transport and diffusion of nanolasers in tissues.

Recently, free-standing miniature lasers, or laser particles (LPs), have emerged as promising optical probes for imaging, sensing, and mass-scale cellular tagging. The spectrally narrowband stimulated emission from LPs can constitute several hundreds of frequency (color) channels, providing much enhanced multiplexing capability compared to conventional fluorescent probes. The multicolor feature makes LPs suited for cell-tracking analysis. III–V compound semiconductors are an attractive choice of materials for miniature lasers due to their high refractive index and high optical gain, as well as good material stability in aqueous media when coated with a protective silica layer.

Nonetheless, semiconductor nanolasers reported to date have two limitations. First, the smallest size of a room-temperature laser has been larger than the optical wavelength in air in at least one dimension, to provide net intracavity gain. The exceptions are resonant Mie scattering nanoparticles made of CsPbBr$_3$ perovskite crystals and spaser (surface plasmon amplification by stimulated emission of radiation) particles involving microbubble cavities. However, the water solubility of the perovskites and the excessive heating of the spasers make them not suited for nonperturbative biological applications. Second, the fabrication of the nanostructures has almost exclusively required electron beam lithography for III–V semiconductors or chemical growth of individual crystals such as for GaN nanowires and II–VI CdS/CdSe nanowires or plates. The electron beam exposure process is time-consuming and expensive, and thus is not acceptable for high-volume production of LPs for barcoding applications that require a large number of LPs. Crystal growth has the potential to meet production-scale requirements but is less established than nanofabrication, especially to directly create III–V semiconductor nanostructures that are suited for biological samples.

Here, we report significant advances in the fabrication and miniaturization of semiconductor nanolasers. First, we demonstrate wafer-scale fabrication of circular nanodisks...
made of an InGa(As)P semiconductor using UV stepper lithography and a plasma ashing method. Our scalable fabrication meets the need to produce a large number of nanodisks on a wafer scale with different colors for tagging biological cells. Second, we achieve room-temperature lasing from subwavelength nanolasers-on-pillar with diameters as small as 460 nm (198 nm height) at optical wavelengths around 660 nm. This represents about 40% reduction in size compared to previous InGaP quantum-well disks fabricated by electron-beam lithography.

We further push the size reduction by the help of a gold substrate via surface plasmon polariton and demonstrate lasing from disks with a diameter of 360 nm. Using a combination of two semiconductor gain materials and diameter variation, we demonstrate lasing from a large number of nanodisk LPs over an 80 nm spectral bandwidth. The multicolor nanodisk LPs coated with a biocompatible silica shell and surface functionalized with polyethylenimine (PEI) are used to demonstrate optical barcoding of various biological cells.

RESULTS/DISCUSSION

We used two different epitaxial structures grown on 3 in. GaAs substrates. Figure 1a shows the first design, in which the active gain material is InGaP with a thickness of 178 nm and is sandwiched by 5 nm thick InGaAlP clad layers to reduce surface recombination on the outer surfaces. Additional 5-nm-thick InGaP layers were added to make the outer surface of disks free of aluminum to avoid oxidation-induced degradation in moist or aqueous environments. The second epitaxial wafer design is described later. Figure 1b shows a representative 3 in. epitaxial wafer scalably patterned with the device structures using stepper UV lithography and O₂ plasma ashing. Figure 1c illustrates the fabrication process by a series of scanning electron microscopy (SEM) images. The first step involves stepper UV lithography to obtain 800 nm diameter circular SU8 photoresist patterns (Figure 1c-i). With typically available production-grade lithography (Auto Step 200, 5× reduction, i-line: 365 nm), it is generally difficult to reliably produce circular patterns below 750 nm in diameter.

To solve this problem, we used oxygen (O₂) plasma ashing to reduce the photosensitive material to smaller dimensions (Figure 1c-ii). The O₂ ashing process transforms the SU8 photosensitive material to gaseous carbon oxides, thereby reducing the diameter (and thickness). For a specific plasma power and O₂ pressure/flow rate, the final diameter of the resist is determined by the ashing time. The resist patterns are then used as etch masks for reactive ion etching (RIE) to obtain the mesa structure (Figure 1c-iii). Finally, the resist is removed,
and GaAs sacrificial layers are selectively etched using a dilute piranha solution (H₂SO₄:H₂O₂:H₂O = 1:1:100). Partial etching results in a disk-on-pillar array on the wafer (Figure 1c-iv). Longer etching detaches the disks completely off the wafer for producing LPs. The smallest possible diameter that can be achieved by this method is limited by the thickness reduction of the photoresist through the ashing process, which does not withstand the full RIE etch time once the ashed photoresist thickness is below ~150 nm. Starting with a 600 nm thick and 800 nm diameter resist, we were able to obtain disk diameters down to ~350 nm. Further details of the fabrication process are given in the Methods section and Figures S1–S3.

Figure 2a shows a 460 nm diameter InGaP disk on an ~120 nm diameter GaAs pillar. We used a nanosecond optical parametric oscillator (OPO) tuned to 560 nm and a hyperspectral microscope to measure the lasing properties (Figure S4). Figures 2b–e show the lasing characteristics of a representative 460 nm diameter InGaP disk on a GaAs pillar. A sharp single-mode lasing emission appeared at 660 nm from the disk above a threshold pump fluence of ~1 mJ/cm² (Figure 2b). The line width decreased from 20 nm below threshold to 0.15–0.2 nm after lasing, and with further increase in pump power, it became broadened due to carrier chirping-induced refractive index changes (Figure 2c). The light-in-light-out plot indicates a spontaneous emission factor (β) of 0.01 (Figure 2d). The laser far-field emission showed interference patterns (Figure 2e). Finite-difference time-domain (FDTD) simulation shows that the lasing mode is of the fourth order (m = 4) transverse electric (TE) whispering gallery mode (WGM) (Figure 2f and Figure S5). The quality factor (Q) of the simulated mode was ~430, although the actual value may be somewhat lower.22 We obtained similar lasing thresholds and line widths from slightly larger disks, but no lasing was observed from disks smaller than 460 nm either on pillars or when fully detached and placed on glass substrates.

We tested nanodisk LPs placed on a gold substrate (Figure 3a) with the expectation that coupling with the gold’s surface plasmon polariton may allow lasing of smaller disks.21,23 We have previously shown plasmonic lasing from submicron CsPbBr₂, perovskite crystals on a gold substrate with lasing thresholds lower than the same gain crystals on glass.21 The Purcell enhancement of the plasmonic mode due to its small mode volume increased the gain and spontaneous emission factor enough to overcome the metallic loss.20 Our numerical and FDTD simulations predicted a similar effect for InGaP disks. For example, the dielectric mode of a 360 nm diameter disk in air has a cavity Q factor of only 20 and a moderate Purcell factor of 2.3. By contrast, the same disk on gold has a much higher-order WGM plasmonic mode localized at the semiconductor–gold interface (Figure 3b). Its cavity Q factor of 30 and Purcell factor of 19 are higher than those of the photonic mode without gold. Note S1 refers to the laser rate equations and parameter definitions used for this analysis. To verify the Purcell enhancement experimentally, we performed time-resolved photoluminescence (PL) spectroscopy. PL decay curves were obtained from three different samples: an intact wafer, a 380-nm-sized disk on a pillar, and a disk of a similar size on a gold substrate (Figure 3c). The bulk semiconductor from the full wafer stack exhibited a monotonic exponential decay with a lifetime of 1840 ps. The disk on pillar showed a monoexponential decay but with a shorter lifetime of 623 ps, evidence of a cavity-enhanced Purcell effect. The disk-on-gold sample showed two distinct exponential decays. The first fast decay has a lifetime less than the photodiode’s response time (<35 ps) and is attributed to plasmonic modes strongly

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Figure 3. Surface plasmon polariton enhanced nanolasers. (a) Schematic of a disk-on-gold laser. (b) FDTD simulated electric field profile, |E|, of the plasmonic mode for a 360-nm-diameter InGaP disk on gold. (c) Time-resolved PL decay curves of the bulk material in the wafer (brown), a disk on a pillar (blue), and a disk on gold (yellow). Circular markers, data points. Lines, double-exponential fits. (d, e) Measured output spectra of two representative disks on gold. Insets: top-view SEM images of the lasers. (f) Measured far-field emission profiles below and above the lasing threshold, and simulated far-field emission profile. (g) Measured line width at different pump fluences. (h) Light-in-light-out curve. (i) Wavelength stability of a device at a pump power level twice the lasing threshold.

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coupled with the gold’s plasmons. The second slow decay with a lifetime of 410 ps is thought to be associated with other photonic-like modes weakly interacting with the plasmons. We observed lasing from 360 nm diameter disks on gold, but not on a pillar or glass.

Figures 3d and 3e show the output spectra of two representative plasmonic devices. The threshold pump fluences were 0.9 and 1.5 mJ/cm$^2$, respectively, similar to the smallest (460 nm diameter) lasers on a pillar. The line width above threshold was about 1.5 nm, considerably broader than the all-dielectric lasers but narrower than the PL line width below threshold (Figure 3g). The performance of plasmonic devices was largely insensitive to the roundness of semiconductor particles (Figure S6). Figure 3f shows the far-field emission profiles of a plasmonic nanolaser below and above the lasing threshold. These profiles correspond well with the simulated image. According to the light-in-light-out plot, the spontaneous emission factor ($\beta$) for the plasmonic device is 0.09, which is 9 times larger than that of the dielectric device. This can be attributed to a significant Purcell effect, as shown in Figure 3h. The lasing performance was stable over repeated pumping (Figure 3i).

Next, we fabricated dielectric nanodisk lasers coated with silica to showcase the potential biological applications of our work. To produce LPs over a wide spectral range, we used a multidisk wafer design$^{24}$ consisting of two disk regions separated by GaAs sacrificial layers (Figure 4a), although it is possible to use two separate wafers each with a different gain material. The pillar structure before and with partial etching of the sacrificial layers was imaged by SEM (Figure 4a bottom). The first disk region is identical to the single-InGaP-active-layer wafer described earlier except for a slightly thinner gain region (158 nm). The second disk region is comprised of an InGaAsP core and InGaP clad layers. The InGaAsP-core gain region has a PL peak at 706 nm, $\sim$56 nm red-shifted from the InGaP-core gain region layer. Using the process described earlier, mesa structures with diameters in the range of 500 to 700 nm were produced. After removing the resist, the entire chip was immersed in dilute piranha solution to selectively etch the GaAs sacrificial layers. Following this, through a multistep centrifugation−resuspension process, we finally obtained a suspension of the nanodisks in ethanol. Finally, they were fully coated with a silica shell of $\sim$25 nm thickness. The entire process to produce silica-coated laser particles, including the removal of the disks from the substrate, silica coating, and purification to obtain laser particles in solution, exhibited a yield of better than 10%.

Figure 4b shows the SEM image of a silica-coated LP with a $\sim$650 nm diameter semiconductor core and $\sim$25 nm silica shell. Figure 4c shows the mode resonances of the silica-coated disks (thickness of 178 nm) fully immersed in water in the diameter range from 430 to 710 nm. The TE and TM modes are interlaced. The free spectral range between neighboring modes is 30−50 nm, which is similar to the semiconductor gain bandwidth of 40 nm. Experimentally, all LPs with diameters greater than $\sim$540 nm showed lasing. Furthermore, nearly all LPs emitted a single lasing peak resulting from the gain competition of neighboring modes. The wavelength tuning slopes as a function of diameter are 0.9 to 1.2 nm/nm for the modes with $m = 4, 5$, and 6. Due to the finite evanescent field energy of the modes outside the silica layer, their resonance wavelengths are somewhat sensitive to the refractive index (RI) of the surrounding medium with coefficients of up to 30 nm shift per RIU (see Figure S5b).
For example, a change in the surrounding index from 1.33 to 1.5 would result in a spectral shift of 5.1 nm for the TE mode. The intended variation of the disk diameters resulted in LPs with a continuum of emission peaks spanning over an 80 nm range from 635 to 715 nm. Figure 4d shows a collection of laser spectra from different LPs suspended in water. The threshold of a silica-coated disk in water was found to be 1.3 mJ/cm², with a spontaneous emission factor of 0.003 and a line width of 0.46 nm (Figure S7). Figure 4e shows the wavelength histograms of LPs from the two individual disk regions, obtained by controlled wet etching to harvest LPs from each disk region separately. The lasing wavelength of the laser particles was stable over an extended operation in water (Figure 4f).

We tested silica-coated LPs for cell tagging after surface functionalization with PEI for efficient uptake. Figure 4g shows RAW 264.7 macrophages, MCF-7 breast cancer cells, and Jurkat T cells in culture, after overnight incubation with LPs. Since the uptake process is stochastic, each cell can internalize a different number of LPs. The presence of LPs in the cytoplasm has no statistically significant effect on the viability of the cells over 72 h (Figure S8), and the cells continue to proliferate and thrive with natural cadence. Figure 4h demonstrates lasing of these LPs from within the cells. The set of LPs associated with each cell acts as their optical barcode, which can be used to distinctly identify individual cells and track them spatiotemporally for large-scale single-cell analysis.²⁰¹¹

CONCLUSIONS

Figure 5 compares our results with those of previously reported dielectric (semiconductor and no metal) and plasmonic (semiconductor and metal) nanolasers. The bulk core-clad structure of our semiconductor nanolasers provided sufficient optical gain to enable InGaP nanolasers as small as 460 nm in diameter. To the best of our knowledge, this represents the smallest III–V semiconductor laser demonstrated to date and the smallest WGM laser made of any materials. Furthermore, the plasmonic lasing achieved with InGaP disks on gold of 360 nm diameter and 198 nm height (~0.02 μm³ in volume) is, in terms of semiconductor volume, the smallest plasmonic laser ever demonstrated at room temperature and is comparable to other nanolasers operated at cryogenic temperatures (Note S2). It should also be noted that this work represents the first optical lithography-based fabrication of submicron-sized LPs. Our results show the potential of multicolor nanodisk LPs in the silicon-detector range for large-scale optical barcoding of cells.

STUDY LIMITATIONS

The present plasmonic lasers require a metal substrate, which limits their biomedical applications. However, it is possible to deposit gold directly onto semiconductor disks on a pillar using electron-beam evaporation to fabricate gold-on-disk structures and eventually to produce standalone plasmonic LPs. Alternatively, it may be possible to coat a gold layer on or around semiconductor disks using redox chemistry. Furthermore, this study explored only two In₀.₅₁Ga₀.₄₉As₀.₅₉P₁₋ₓ stoichiometries that spanned an 80 nm wavelength range. It should be possible to extend the spectral range to a wavelength of 1000 nm, for example, by incorporating different semiconductor compositions. The operating pump levels of a few mJ/cm² are substantially higher than those of other low-threshold laser devices. It corresponds to several pJ per pump pulse. However, this excitation fluence is thought to be adequate for biological applications where the pump beam is rapidly scanned over the sample and could be within the laser safety limit of biological tissues (~0.25 J/cm² for an exposure time of 2 seconds in the skin).²⁵ For example, pulse repetition rates of up to 100 kHz would be acceptable for an exposure time of 10⁻³ s. Finally, the sensitivity of the lasing wavelength to the surrounding medium may be an issue when such spectral shifts are not desired. We note that the sensitivity can be significantly reduced with thicker silica coating (e.g., >50 nm). On the other hand, most cell tagging applications can tolerate certain wavelength shifts. We found that silica-coated InGaAsP LPs, once incorporated into the cytoplasm or attached on the cell membrane, remain stable within a wavelength resolution of ±0.5 nm. We have used this spectral barcoding strategy in our recent imaging and flow cytometry studies.²⁰¹¹ Compared to the previous LPs in the spectral range of NIR-II (1100–1600 nm), the LPs developed in this work offer smaller sizes and compatibility with silicon detector technology. However, their spectral range overlaps with some commonly used dyes in the red and NIR-I ranges, and therefore, it limits the compatibility with fluorescence techniques.

METHODS/EXPERIMENTAL

Epitaxial Wafer Structure. The devices are fabricated from two different wafer structures. In both cases, the substrate is GaAs. In the first wafer structure, only a single active layer material is present, with the exact structure being GaAs substrate/600 nm GaAs/5 nm InGaP/5 nm In₀.₄₉Ga₀.₅₁As₀.₅₀P₀.₅₀P/178 nm In₀.₅₁Ga₀.₄₉P/5 nm In₀.₄₉Ga₀.₅₁As₀.₃₁P₀.₆₉P/5 nm In₀.₃₈Ga₀.₆₂As₀.₃₁P₀.₆₉P/5 nm In₀.₃₈Ga₀.₆₂As₀.₃₁P₀.₆₉P/300 nm GaAs. The central InGaP forms the main active material layer, while the 10 nm layers on either side form a cladding layer to reduce surface recombination. The top 300 nm GaAs is a sacrificial capping layer to protect the active material from potential damage during fabrication. In the second wafer structure, two active layer materials are present, with the exact structure being GaAs substrate/400 nm GaAs/10 nm InGaP/170 nm InGaAsP/10 nm InGaP/400 nm GaAs/5 nm In₀.₅₁Ga₀.₄₉P/158 nm In₀.₅₁Ga₀.₄₉P/5 nm In₀.₄₉Ga₀.₅₁As₀.₃₁P₀.₆₉P/5 nm In₀.₃₈Ga₀.₆₂As₀.₃₁P₀.₆₉P/300 nm GaAs. GaAs layers are sacrificial. These are
custom ordered and epitaxially grown using metal organic chemical vapor deposition (MOCVD) by Xiamen Powerway Advanced Materials Co. Ltd.

**Fabrication of Microdisk Lasers on a Chip.** Device fabrication starts with cleaning the wafers in acetone, isopropyl alcohol (IPA), methanol, and deionized water to remove dissolvable contaminants from the surface. The wafer is baked at 180 °C to dehydrate adsorbed water molecules, and then processed by oxygen plasma to desumb and remove any remaining photoresist molecules on the unexposed substrate surface. To get descummed using oxygen plasma to remove any remaining photoresist, a mixture of 10:1:100 is used to selectively etch the GaAs layers while keeping the active materials intact. Partial etching of the GaAs layers creates a pillar structure with the diameter smaller than the overlying disk, thereby creating the nanodisk structures on a chip.

**Fabrication of Free-Standing Laser Particles.** After the nanodisk structures on the chip are created, complete etching of GaAs releases the nanodisks in the piranha solution. The next steps involve collection of the laser particles and making them biocompatible for cell studies. The laser particle suspension in piranha is centrifuged to collect the disks at the bottom of a tube, and the supernatant liquid is selectively removed. Ethanol is then added to the tube, and the laser particles are redispersed in it by ultrasonication. Repeating this centrifugation, decanting of supernatant, adding fresh ethanol, and resuspension of laser particles, the medium is effectively cleared of piranha solution. Using tetraethoxysilicate (TEOS) with the reaction initiated in a basic medium (through ammonium hydroxide), a uniform silica shell is then created around each disk to obtain freely suspended biocompatible silica-coated laser particles. These are then surface-functionalized with PEI for improved cellular association.

**Optical Characterization.** For the laser experiment, the specimen is placed in a home-built epi-fluorescence microscopy setup. The pump source is an optical parametric oscillator (Optotek HE 355 LD) tuned to 650 nm, with a repetition rate of 20 Hz and a pulse duration of 4 ns, or a frequency-doubled NdYAG laser (532 nm), with a repetition rate of 1 kHz and a pulse duration of 4 ns. Using a 0.6 NA, 50X air objective lens or 0.9 NA, 100X air objective lens (Nikon), the full-width-at-half-maxima (fwhm) size of the pump beam on the sample is about 20 μm. The emission from the sample collected by the objective lens is passed through a dichroic mirror and a dichroic filter and split to a silicon-based EMCCD camera ( Luca, Andor) for wide-field imaging and to a grating-based EMCCD spectrometer (Shamrock, Andor). With an entrance slit width of 20 μm, the measurement spectral resolution is about 0.13 nm. For time-resolved photoluminescence measurements, a picosecond laser (VisIR-765, PicoQuant) is used, which is frequency doubled to 382 nm using a nonlinear Beta barium borate (BBO) crystal, a single-photon avalanche photodiode (MicroPhotonics Devices) with a single photon timing resolution of 35 ps (fwhm), and a time-correlated single-photon counting board (TimeHarp 260, PicoQuant).

**Fabrication of the Plasmonic Laser.** The free-standing disks suspended in ethanol solution are simply drop-cast on a polycrystalline gold substrate (Platypos). The root-mean-square roughness of the gold surface is about 0.7 nm. After characterizing the lasing properties of the device, we mark the corresponding device’s position by irradiating the gold substrate using high pump fluence (Figure S10). The devices are then imaged by SEM to precisely quantify their sizes.

**Finite Difference Time Domain Simulation.** The eigenmodes and field profiles of the nanodisk lasers are numerically simulated with a commercial 3D FDTD solver from Lumerical. The optical constants for gold are taken from the CRC handbook. In order to excite the desired modes, an electric dipole is positioned inside the semiconductor. The 3D simulation utilizes a mesh grid size of 5 nm or less with mesh refinement by the Yu–Mitra method. The boundary conditions are implemented by using perfectly matched layers. To measure the time-dependent electric fields, point-like time monitors are placed inside or near the semiconductor particle. The recorded data are analyzed in the spectral domain by a fast Fourier transform. The cavity Q factor is calculated from $\sigma_{res}/\Delta\nu_{res}$ using the peak wavelength position $\sigma_{res}$ and the full width half-maximum $\Delta\nu_{res}$. To obtain a 3D near-field pattern of the modes, we utilize an array of 2D field monitors covering the entire simulation volume with time apodization. To avoid the effect of transients, the apodization center is set to 420 fs with a width of 100 fs.

**Cell Experiment.** HeLa human cervical cancer cells, MCF-7 human breast cancer cells, RAW 264.7 mouse macrophage cells, and Jurkat Clone E6-1 human T lymphoblast (Jurkat T) cells are purchased from ATCC (American Type Culture Collection). HeLa, MCF-7, and RAW 264.7 cells are cultured and maintained in the cell media of Dulbecco’s modified Eagle medium (DMEM) supplemented with 10% (v/v) fetal bovine serum (FBS) and 1% (v/v) penicillin–streptomycin at 37 °C under 5% CO2. Jurkat T cells are cultured in RPMI 1640 (Thermo Fisher) cell medium supplemented with 10% FBS and 1% antibiotic–antimycotic. For cytotoxicity assessment, HeLa and RAW 264.7 cells are seeded in 96-well plastic plates with the density of 3000 cells/well and incubated first for 24 h. Then about 6000 silica-coated visible LPs are added into each well. After further incubation for 24, 48, and 72 h, the standard cell counting kit (CCK8) (Sigma-Aldrich) assay is performed to test the cell viability. The nontreated cells at different time points are used as control (100%) for calculation. For each group, six parallel experiments are performed simultaneously. For the cell tagging experiment, cells are plated in their respective media at a known density on a glass-bottom plate. After 24 h of incubation, the visible laser particles resuspended in PBS buffer are added into cells with the particle-to-cell ratio 4:1. The plate is shaken to mix the media evenly, and the cell media is carefully refreshed 6 h later. After 24 h of further incubation, the cells are washed three times with DPBS buffer and fixed using a 4% paraformaldehyde (PFA) solution for the following confocal imaging (Olympus, FV3000) and cell laser experiment.

**ASSOCIATED CONTENT**

**Supporting Information**

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.nano.3c04721.

SEM images of fabrication of 800 nm diameter disks; top view SEM images of disks-on-pillar; SEM images showing geometry control of photoresist by O2 plasma ashing; schematic of laser optical characterization setup; FDTD simulation results of mode resonance; sideways SEM image of a 350 nm diameter disk and plane-view SEM images of irregular shaped disks and their output emission spectra; lasing characteristics of silica-coated InGaP disk in water; cell viability analysis of RAW 264.7 macrophage and HeLa cells; effect of different reactive ion etching processes on disk morphology; method for identifying individual disks on a gold substrate under...
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Author Contributions
DS and SC contributed equally to this work. DS, SC, and SHY designed the project. DS developed and performed the nanofabrication. SC performed optical measurements and computer simulations. HY conducted the cell culture. NM and PHD contributed to the nanofabrication and measurement. DS, SC, and SHY wrote the manuscript with input from all authors.

Notes
The authors declare the following competing financial interest(s): PHD, NM, and SHY hold patents on laser particle technologies. The financial interests of NM and SHY were reviewed and are managed by Mass General Brigham in accordance with their conflict-of-interest policies.

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ABBREVIATIONS
LP, laser particle; SEM, scanning electron microscopy; WGM, whispering gallery mode; FDTD, finite-difference time-domain

REFERENCES
(2) Liang, Y.; Li, C.; Huang, Y.-Z.; Zhang, Q. Plasmonic Nanolasers in On-Chip Light Sources: Prospects and Challenges. ACS Nano 2020, 14 (11), 14375–14390.
(13) Tigunset, E.; Koshelev, K.; Furasova, A.; Tonkaev, P.; Mikhailovskii, V.; Ushakova, E. V.; Baranov, D. G.; Shegai, T.; Zakhidov, A. A.; Kivshar, Y.; Makarov, S. V. Room-Temperature


